PROCEEDINGS OF UNIVERSITIES. APPLIED CHEMISTRY AND BIOTECHNOLOGY 2021 Vol. 11 No. 4 ИЗВЕСТИЯ ВУЗОВ. ПРИКЛАДНАЯ ХИМИЯ И БИОТЕХНОЛОГИЯ 2021 Том 11 N 4

PHYSICOCHEMICAL BIOLOGY

Original article DOI: https://doi.org/10.21285/2227-2925-2021-11-4-531-539



Synthesis and crystal structure of cytotoxic copper(II) complex with 1,10-phenanthroline-5,6-dione and isothiazole derivative

Yuliya A. Golubeva*, Ksenia S. Smirnova*, Lubov' S. Klyushova**, Vladimir I. Potkin***, Elizaveta V. Lider*

*Nikolaev Institute of Inorganic Chemistry, SB RAS, Novosibirsk, Russian Federation **Institute of Molecular Biology and Biophysics, Federal Research Center foe Fundamental and Translational Medicine, Novosibirsk, Russian Federation ***Institute of Physical Organic Chemistry of the National Academy of Sciences of Belarus, Minsk, Belarus

Corresponding author: Elizaveta V. Lider, lisalider@ngs.ru

Abstract. Oligopyridine based copper(II) complexes are of interest to scientists as possible anticancer agents due to promising cytotoxic and DNA binding/cleaving properties. In this study, copper(II) complex [Cu(phendione)L₂]·C₂H₅OH with 1,10-phenanthroline-5,6-dione (phendione) and 4,5-dichloro-isothiazole-3carboxylic acid (HL) was synthesized and characterized by elemental analysis, IR-spectroscopy, X-ray powder diffraction and single-crystal X-ray diffraction. According to X-ray diffraction data, obtained compound is mononuclear complex with square pyramidal coordination environment of the central atom which is surrounded by two isothiazolate molecules and one phendione ligand. The X-ray diffraction data are confirmed by IR-spectroscopy data showing the presence of characteristic stretching vibration bands of the carbonyl and carboxyl groups of oligopyridine ligand and isothiazolate ions, respectively. Density functional theory (DFT) calculations for complex were carried out using the ADF software package to perform geometry optimization and frequency calculations that were in a good agreement with experimental IR spectrum. Cytotoxicity of complex and initial reagents was tested in vitro against HepG2 (human hepatocellular carcinoma) and MCF-7 (human breast adenocarcinoma) cell lines. The complex showed high dose-dependent cytotoxic activity with the IC₅₀ values of 0.60±0.03 μ M and 0.96±0.13 μ M, respectively, which is higher than the activity of cisplatin against these cell lines. The activity of the complex is due to the presence of phendione ligand, which exhibits a similar cytotoxic activity.

Key words: copper complex, isothiazole, crystal structure, phenanthroline, cytotoxicity

Acknowledgments. The authors thank A. P. Zubareva and N. N. Komardina for the elemental analysis, A. O. Matveeva for the X-ray phase analysis data, T. S. Sukhikh for providing the data collected in XRD Facility of NIIC SB RAS. The work was performed using the equipment of the Center for Collective Use "Proteomic Analysis", supported by funding from the Ministry of Science and Higher Education of the Russian Federation (agreement no. 075-15-2021-691).

Funding. The reported study was funded by RFBR (project no. 20-33-90092).

The research (Nikolaev Institute of Inorganic Chemistry SB RAS) was supported by the Ministry of Science and Higher Education of the Russian Federation (no. 121031700321-3).

For citation: Golubeva Yu. A., Smirnova K. S., Klyushova L. S., Potkin V. I., Lider E. V. Synthesis and crystal structure of cytotoxic copper(II) complex with 1,10-phenanthroline-5,6-dione and isothiazole derivative. *Izvestiya Vuzov. Prikladnaya Khimiya i Biotekhnologiya = Proceedings of Universities. Applied Chemistry and Biotechnology.* 2021;11(4):531-539. (In English). https://doi.org/10.21285/2227-2925-2021-11-4-531-539.

[©] Golubeva Yu. A., Smirnova K. S., Klyushova L. S., Potkin V. I., Lider E. V., 2021

ФИЗИКО-ХИМИЧЕСКАЯ БИОЛОГИЯ

Научная статья УДК 54.057;546.562;615.277.3

Синтез и кристаллическая структура цитотоксического комплекса меди(II) с 1,10-фенантролин-5,6-дионом и производным изотиазола

Юлия Андреевна Голубева*, Ксения Сергеевна Смирнова *, Любовь Сергеевна Клюшова **, Владимир Иванович Поткин ***, Елизавета Викторовна Лидер *

*Институт неорганической химии им. А. В. Николаева СО РАН,

г. Новосибирск, Российская Федерация

**Научно-исследовательский институт молекулярной биологии и биофизики – структурное подразделение Федерального исследовательского центра фундаментальной и трансляционной медицины, г. Новосибирск, Российская Федерация

***Институт физико-органической химии Национальной академии наук Беларуси,

г. Минск, Беларусь

Автор, ответственный за переписку: Лидер Елизавета Викторовна, lisalider@ngs.ru

Аннотация. Комплексы меди(II) на основе олигопиридинов привлекают интерес исследователей в качестве возможных противоопухолевых агентов ввиду их выдающихся цитотоксических свойств и способности связываться/расшеплять ДНК. В настоящем исследовании был получен и охарактеризован с помощью ИК-спектроскопии, элементного, рентгенофазового и рентгеноструктурного анализа комплекс меди(II) с 1,10-фенантролин-5,6-дионом (phendione) и 4,5-дихлоро-изотиазол-3-карбоновой кислотой, [Cu(phendione)L₂] C_2H_5OH . Согласно данным рентгеноструктурного анализа, полученное соединение является моноядерным, при этом две молекулы изотиазола и 1,10-фенантролин-5,6-дион образуют квадратно-пирамидальное окружение центрального атома. Данные рентгеноструктурного анализа согласуются с данными ИК-спектроскопии, указывающими на наличие характеристичных полос валентных колебаний карбонильной и карбоксильной групп олигопиридина и производного изотиазола соответственно. Расчеты, выполненные методом теории функционала плотности (DFT) с использованием программного пакета ADF, позволили оптимизировать геометрию комплекса и вычислить теоретический ИК-спектр, который хорошо согласуется с экспериментальным. Цитотоксичность комплексов и исходных реагентов исследована на клеточных линиях HepG2 (гепатоцеллюлярная карцинома) и MCF-7 (аденокарцинома молочной железы). Комплекс проявляет высокую дозозависимую цитотоксическую активность, при этом значения параметра IC₅₀ составляют 0,60±0,03 мкМ (НерG2) и 0,96±0,13 мкМ (МСF-7), что превышает активность цисплатина по отношению к данным клеточным линиям. Активность комплекса обусловлена присутствием лиганда phendione, который в свободном виде также обладает токсичностью.

Ключевые слова: комплекс меди, изотиазол, кристаллическая структура, фенантролин, цитотоксичнсоть

Благодарности. Авторы выражают благодарность А. П. Зубаревой и Н. Н. Комардиной за предоставление данных элементного анализа, А. О. Матвеевой – за предоставление данных рентгенофазового анализа, Т. С. Сухих – за предоставление данных, измеренных в рентгенодифракционном ЦКП ИНХ СО РАН.

Финансирование. Исследование выполнено при финансовой поддержке РФФИ в рамках научного проекта № 20-33-90092.

Работа выполнена с использованием оборудования ЦКП «Протеомный анализ», поддержанного финансированием Минобрнауки России (соглашение № 075-15-2021-691).

Для цитирования: Голубева Ю. А., Смирнова К. С., Клюшова Л. С., Поткин В. И., Лидер Е. В. Синтез и кристаллическая структура цитотоксического комплекса меди(II) с 1,10-фенантролин-5,6-дионом и производным изотиазола // Известия вузов. Прикладная химия и биотехнология. 2021. Т. 11. N 4. С. 531–539. https://doi.org/10.21285/2227-2925-2021-11-4-531-539.

INTRODUCTION

Currently, mixed-ligand complexes of essential metals with 1,10-phenanthroline and its derivatives are being actively studied in vitro and in vivo for their biological properties (cytotoxic, antioxidant, antibacterial, antiviral and other activities) [1-7]. Essential metals include such microelemets as iron, copper, zinc, cobalt, nickel, manganese and some other metals. Among the complexes of essential metals, the most interesting as anticancer agents are 1,10-phenanthroline based copper(II) complexes, since they are capable to interact and cleave DNA/RNA backbone [8, 9]. Moreover, oligopyridine and amino acid/acetylacetone based copper(II) complexes (Casiopeínas) have shown antiproliferative, genotoxic, and antineoplastic activity [10, 11]. The most promising complexes from the Casiopeinas series have completed preclinical trials [12]. Besides amino acids, N-donor systems (terpyridine, imidazole, benzimidazole, tetrazole), O-donor systems (salicylic acid, dicarboxylic acids), S-donor systems (thiosemicarbazones, dithiocarbamates, thioureas), Schiff bases are often used as secondary ligand in synthesis of cytotoxic Casiopeinas-like complexes [6]. The isothiazole heterocycles and coordination compounds with isothiazoles have been shown to be bioactive substances with pesticidal, anticancer, anti-inflammatory and antiviral activities [13]. However, despite their promising properties, isothiazoles are negligibly used for the synthesis of bioactive oligopyridine based complexes with essential metals. Thus, the research aiming preparation of corresponding complexes may be indeed promising from the point of view of medicinal chemistry.

In continuation of our work [14] in order to expand the number of available cytotoxic oligopyridine based complexes with isothiazole as secondary ligand, the present study set out to synthesise new mixed-ligand copper(II) complex with 1,10-phena-nthroline-5,6-dione and 4,5-dichloro-isothiazole-3-carboxylic acid and evaluate its biological activity.

EXPERIMENTAL SECTION

1,10-Phenanthroline-5,6-dione (phendione) was acquired from ABCR (Germany). 4,5-Dichloro-isothiazole-3-carboxylic acid (HL) was synthesized as previously described [15, 16]. Solvents and reagents were used as purchased without any further purification.

Elemental analysis (C, H, N) was performed using Euro EA 3000 analyzer. IR absorption spectra were recorded on SCIMITAR FTS 2000 and VERTEX-80 spectrophotometers at 4000–400 cm⁻¹ (in fluorinated oil – in the region of 4000–1500 cm⁻¹, in vaseline oil – 1800–400 cm⁻¹). Powder XRD analysis of complexes was performed on a Shimadzu XRD-7000 diffractometer (CuK α radiation, Ni filter, 3–40° 20 range, 0.03° 20 step, room temperature). Quantum-chemical calculations were carried out on the computational cluster of NIIC SB RAS using the Amsterdam Density Functional (ADF) program by dint of density functional theory (DFT)¹. The generalized gradient approximation (GGA), the density functional PBE (Perdew – Burke – Ernzerhof) [17] in combination with the all electron basis set TZ2P [18] were applied. The calculations were carried out for non-solvent system.

Bruker D8 Venture diffractometer with the graphite-monochromated MoK_a radiation ($\lambda = 0.71073$ Å) was utilized to collect single-crystal XRD data for copper(II) complex. All measurements were carried out at 150 K, and the $\phi\text{-}$ and $\omega\text{-}\text{scan}$ techniques were employed. Absorption corrections were applied with the SADABS program². The crystal structure was solved and refined by means of the SHELXT [19] and SHELXL [20] programs using OLEX2 GUI [21]. Atomic thermal displacement parameters for non-hydrogen atoms were refined anisotropically. Positions of hydrogen atoms were calculated according to their geometrical conditions and refined by dint of the riding model. The crystallographic data and details of the structure refinements are shown in Table 1. CCDC 2121029 contains the supplementary crystallographic data for this paper. These information can be obtained free of charge from The Cambridge Crystallographic Data Center at https://www.ccdc.cam.ac.uk/structures/.

Synthesis of [Cu(phendione)L₂]·C₂H₅OH. [Cu(H₂O)L₂] was synthesized as previously described [14]. The precipitate of [Cu(H₂O)L₂] (0.048 g, 0.10 mmol) was resuspended in mixture of ethanol (2 ml) and dichloromethane (3 ml). Solid phendione (0.025 g, 0.12 mmol) was added to suspension with stirring. The reaction mixture became clear, precipitate dissolved. The solution was left for slow evaporation at room temperature. Polycrystalline phase was obtained after a week. Green crystals were filtered out, washed with ethanol and dried in air. Yield: 0.064 g (90%). Elemental analysis (%): Calc. for $C_{22}H_{12}Cl_4CuN_4O_7S_2$: C, 37.0; H, 1.7; N, 7.8. Found: C, 37.3; H, 1.8; N, 7.7.

Cytotoxic activity. Human hepatocellular carcinoma (HepG2) and human breast adenocarcinoma (MCF-7) cell lines were cultured in Iscove's Modified Dulbecco's Medium (IMDM) supplemented with a 10% fetal bovine serum under a humidified atmosphere (5% CO₂ and 95% air) at 37 °C. Cell viability was evaluated by Hoechst/PI staining as previously described [22]. Hep2 cells were cultured to 96-well plates at a density 5 $\cdot 10^3$ cells per well. After 24 hours cells were treated with complex dissolved in DMSO and incubated for 48 hours. Serial dilutions were prepared in IMDM medium in the concentration range of 0.1–25 µM. For identification of live, apoptotic and dead cells, treated cells and control cells were stained with a mixture of fluorescent dyes

¹ADF2013, Software for Chemistry & Materials, Theoretical Chemistry, Vrije Universiteit, Amsterdam (The Netherlands), 2013. ²Bruker Apex3 software suite: Apex3, SADABS-2016/2 and SAINT, version 2018.7-2; Bruker AXS Inc.: Madison, WI, 2017.

Hoechst 33342 (Sigma-Aldrich) and propidium iodide (Invitrogen) for 30 min at 37 °C. An IN Cell Analyzer 2200 (GE Healthcare, UK) was used to perform automatic imaging of four fields per well under 200×magnification, in bright field and fluorescence channels. The IN Cell Investigator image analysis software (GE Healthcare, UK) was used to determine live, apoptotic and dead cells among the whole population. All data shown are mean of three wells. The quantitative data were expressed as the mean ± standard deviation (SD). All statistical analyses were performed using the software Excel 2016 (Microsoft) and Origin 8.0.

 Table 1. Crystallographic data

 of the copper(II) complex

Таблица 1. Кристаллографические данные для комплекса меди(II))

Parameter	Value
Empirical formula	$C_{22}H_{12}CI_4N_4O_7S_2Cu$
Formula weight	713.82
Crystal system,	Monoclinic, P21/c
space group	
lemperature (K)	150
a/A	11.2744(15)
	8.5017(11)
	20.401(3)
ß/°	94 024(5)
2/ ⁰	90
Volume/Å ³	2723 2(6)
Z	4
$\rho_{calc} g/cm^3$	1.741
µ/mm ⁻¹	1.399
Crystal size/mm	0.3 × 0.2 × 0.12
2O range	4 774 to 51 486
for data collection/°	
Index ranges	$-13 \le h \le 7, -10 \le k \le 10,$
	-33 ≤ 1 ≤ 34
Reflections collected	34013
Independent reflections	$5030 [R_{int} = 0.0520,$
Restraints/parameters	$R_{sigma} = 0.0390$
$Goodness-of-fit on F^2$	1 088
Final R indexes $[l>=2\sigma(l)]$	$R_1 = 0.0423$, $wR_2 = 0.0921$
Final R indexes [all data]	$R_1 = 0.0554$, $wR_2 = 0.0971$
Largest diff.	0.75/ 0.41
peak/hole / e/Å ⁻³	0.75/-0.41

RESULTS AND DISCUSSION

Green crystals of copper(II) complex $[Cu(phendione)L_2] \cdot C_2H_5OH$ were obtained after slow evaporation of ethanol/dichloromethane (1:1.5 by volume) solution containing $[Cu(H_2O)L_2]$ and phendione ligand in 1:1.2 molar ratio. Obtained complex is soluble in DMSO, CH₃CN, sparingly soluble in ethanol, CH₂Cl₂ and practically insoluble in water. The elemental analysis results for $[Cu(phendione)L_2] \cdot C_2H_5OH$ are consistent with the proposed formula of the complex. The presence of solvent ethanol molecule was confirmed by IR-spectroscopy (v(OH) = 3464, 3308 cm^{-1}). Powder X-ray diffraction analysis have been used to demonstrate the identity of single crystal and the synthesized polycrystalline phase of complex which proved the phase purity of the bulk sample (Fig. 1).

According to single-crystal X-ray diffraction data, the copper(II) complex [Cu(phendione)L₂]·C₂H₅OH reveals square pyramidal coordination environment of central atom. The coordination sphere of Cu(II) consists of two O atoms of the isothiazoles carboxylates, one N of the isothiazole ligand and two N atoms of the phendione (Fig. 2). Additional contact (of 2.772 Å) is observed between the copper(II) and the O atom of the isothiazole moiety giving distorted octahedral 5 + 1 environment. Also there is outer-sphere ethanol molecule. Similar structure has been observed in our previous work, where bipicoline has been used instead of phendione [14].



Fig. 1. X-ray powder patterns for $[Cu(phendione)L_2] \cdot C_2H_5OH$ **Рис. 1**. Дифрактограмма для $[Cu(phendione)L_2] \cdot C_2H_5OH)$



Fig. 2. Structure of $[\text{Cu}(\text{phendione})L_2]$ complex (Solvent ethanol molecule is not shown)

Рис. 2. Структура комплекса [Cu(phendione)L₂] (молекула растворителя не показана)

DFT-calculations, such as IR-frequencies and energies of HOMO/LUMO, have been carried out for copper(II) complex. The highest occupied molecular orbital (HOMO) and lowest-lying unoccupied molecular orbital (LUMO) are known to be the most im-

portant orbitals in a molecule. In case of copper(II) complex, the electron density of HOMO (Fig. 3) is concentrated on phendione mainly and energy of this orbital is equal to -5.978 eV. LUMO (-4,997 eV) is disposed on copper ion and donor atoms of its coordination sphere. The energy gap between HOMO and LUMO ($E_{HOMO} - E_{LUMO}$) is -0.981 eV. A molecule with large HOMO–LUMO gap is described as a hard molecule which is much less polarizable. In contrast, the soft systems have small HOMO–LUMO gap and are highly polarizable [23]. Thus, complex refers to soft molecules due to the small value of energy gap.

Vibrational spectrum has been calculated for the complex with optimized geometry. There are no imaginary frequencies in the obtained spectrum, consequently, the geometry corresponds to local minima. The main vibrational frequencies of calculated and experimental spectra are shown in Table 2. The vibrations of characteristic groups (COO-, C-H, C-CI, O=C-C=O) exhibit in both spectra. The slight shift and broadening of bands is observed in experimental IR-spectrum, which can be due to the presence of ethanol molecule in system. Despite these small differences spectra are in a good agreement.

The obtained IR-spectrum clearly indicates the formation of copper(II) complex. There is a disappearing of broad v(O–H) band observed in the HL spectrum (2915 cm⁻¹) in the result of ligand deprotonation during the synthesis. Moreover, the bands of asymmetrical stretching vibrations of the carboxylate group shift to the low-frequency region (1651–1614 cm⁻¹) in contrast to the band in HL spectrum (v(COO)_{as} = 1724 cm⁻¹).

The cytotoxicity of copper(II) complex was tested by Hoechst/PI staining on HepG2 (human hepatocellular carcinoma) and MCF-7 (human breast adenocarcinoma) cell lines by exposing them for 48 h to the medium containing the compound in 0.1–25 μ M concentration. The concentration-dependent cell viability graphs are given in Fig. 4. Calculated from these graphs IC₅₀ values (concentration required to reduce survival in the cell lines to 50%) are summarized in Table 3.

Earlier, copper acetate and HL were shown to be nontoxic in the concentration range of $1-50 \mu$ M [14]. At the same time, 1,10-phenanthroline-5,6-dione is highly toxic to HepG2 and MCF-7 cells with IC₅₀ values being in the nanomolar concentration range [24].



Рис. 3. Электронная плотность [Cu(phendione) L_2] (a), B3MO (b) и HCMO (c)

Table 2. Calculated and experimental vibrational frequencies (cm^{-1}) of [Cu(phendione)L₂]

Таблица 2. Теоретически рассчитанные и экспериментальные колебательные частоты (см ⁻¹)
комплекса [Cu(phendione)L ₂]	

Assignment, cm ⁻¹	DFT-calculation	Experimental data
ν(O–H) _{EtOH}	_	3464, 3308
ν(C–H)	3130, 3115, 3106	3101, 3086, 3061
v(C–H) _{EtOH}	_	2960, 2924, 2856
v(O=C-C=O)as	1692	1726
$v(O=C-C=O)_s$	1664	1710
v(COO) _{as}	1642, 1628	1651, 1614
v(COO)s	1261	1261
v(C–Cl)	1055, 1044	1049, 1024
$R_{rings}, \delta(C-H)$	1578, 1560, 1412, 1306, 1195, 1114, 979, 942	1576, 1408, 1300, 1205, 1128, 972



Fig. 4. Effect of [Cu(phendione)L₂]·C₂H₅OH on the viability of HepG2 and MCF-7 cells

Рис. 4. Эффект воздействия [Cu(phendione)L₂]·C₂H₅OH на выживаемость клеток HepG2 и MCF-7

According to cytotoxicity study of [Cu(phendione)L₂]·C₂H₅OH, complex possesses dose-dependent cytotoxicity against both cell lines (IC₅₀ = 0.96±0.13 μ M (MCF-7), 0.60±0.03 μ M (HepG2)), while its toxicity is higher than that of oligopyridine on HepG2 cells and vice versa on MCF-7 cells (See Table 3). Thus, the activity of the complex is due to the presence of phendione ligand, which exhibits a similar cytotoxic activity. The activity of the obtained complex exceeds the

activity of cisplatin by more than an order of magnitude.

This compound is part of a series of mixed-ligand copper(II) complexes with HL and various oligopyridines, and turned out to be the most toxic in this series. So, for example, the IC₅₀ value against MCF-7 cells in the same conditions is $1.8\pm0.3 \ \mu$ M for [Cu(4,7-dimethyl-1,10-phenanthroline)L₂] and $4.2\pm0.2 \ \mu$ M for [Cu(1,10-phenanthroline)(H₂O)L₂] [14].

Table 3. IC_{\rm 50} values of the compounds against MCF-7 and HepG2 cell lines

Таблица 3. Значения IC₅₀ для полученных соединений по отношению к клеточным линиям MCF-7 и HepG2

Compound	IC ₅₀ value, μΜ		
Compound	MCF-7 cells	HepG2 cells	
Cu(OAc) ₂	>50	>50	
HL	>50	_	
1,10-phenanthroline-5,6-dione	0.73±0.06	3.3±0.6	
[Cu(phendione)L₂]·C₂H₅OH	0.96±0.13	0.60±0.03	
Cisplatin	33.7±1.8	33.0±5.4	

CONCLUSIONS

Mononuclear copper(II) complex based on 4,5-dichloro-isothiazole-3-carboxylic acid and 1,10-phenanthroline-5,6-dione [Cu(phendione)L₂] has been obtained and characterized by several physicochemical methods. The results of single crystal X-ray diffraction indicate that complex reveals square pyramidal coordination environment of central atom which consist of two O atoms of the isothiazole scarboxylates, one N of the isothiazole ligand and two N atoms of the phendione. According to DFT-calculations, the value of energy gap between HOMO and LUMO of complex is small (-0.981 eV), thus, [Cu(phendione)L₂] refers to soft

molecules. This compound causes pronounced cytotoxic effect against HepG2 and MCF-7 cells in 0.1-25 concentration range which is an order of magnitude higher than cisplatin cytotoxicity. [Cu(phendione)L₂] is part of a series of mixed-ligand copper(II) complexes with HL and various oligopyridines. Cytotoxicity of these complexes increases with the transition from 1,10-phenanthroline to 4,7-dimethyl-1,10-phenanthroline and is maximal in the 1,10-phenanthroline-5,6-dione based complex. Thus, this work helps to understand structure-cytotoxic activity relationships for the oligopy-ridine based isothiazole copper(II) complexes.

REFERENCES

1. Bencini A., Lippolis V. 1 ,10-Phenanthroline: A versatile building block for the construction of ligands for various purposes. *Coordination Chemistry Reviews.* 2010;254(17):2096–2180. https://doi. org/10.1016/j.ccr.2010.04.008.

2. Krasnovskaya O., Naumov A., Guk D., Gorelkin P., Erofeev A., Beloglazkina E., et al. Copper coordination compounds as biologically active agents. International Journal of Molecular Sciences. 2020;21(11): 3965. https://doi.org/10.3390/ijms21113965.

3. Dey D., Roy A. B., Ranjani A., Gayathri L., Chandraleka S., Dhanasekaran D., et al. Synthesis and bio-catalytic activity of isostructural cobalt(III)phenanthroline complexes. *Journal of Chemical Sciences*. 2015;127(4):649–661. https://doi.org/10. 1007/s12039-015-0817-y.

4. Al-Omair M. A. Biochemical activities and electronic spectra of different cobalt phenanthroline complexes. *Arabian Journal of Chemistry*. 2019;12(7):1061–1069. https://doi.org/10.1016/j.arabjc.2018.11.006.

5. Čongrádyová A., Jomová K., Kuckova L., Kožíšek J., Moncol' J., Valko M. Antimicrobial activity of copper(II) complexes. *Journal of Microbiology, Biotechnology and Food Sciences*. 2014;3(1):67–70.

6. Mahalakshmi R., Raman N. A Therapeutic journey of mixed ligand complexes containing 1,10-phenanthroline derivatives: A review. *International Journal of Current Pharmaceutical Research*. 2016;8(3):1–6.

7. Viganor L., Howe O., McCarron P., McCann M., Devereux M. The antibacterial activity of metal complexes containing 1,10-phenanthroline: Potential as alternative therapeutics in the era of antibiotic resistance. *Current Topics in Medicinal Chemistry*. 2016;17(11):1280–1302. https://doi.org/10.2174/15680266166666161003143333.

8. Galindo-Murillo R., García-Ramos J. C., Ruiz-Azuara L., Cheatham T. E., Cortés-Guzmán F. Intercalation processes of copper complexes in DNA. *Nucleic Acids Research.* 2015;43(11):5364–5376. https://doi.org/10.1093/nar/gkv467.

9. Sigman D. S., Graham D. R., D'Aurora V., Stern A. M. Oxygen-dependent cleavage of DNA by the 1,10-phenanthroline.cuprous complex. Inhibition of *Escherichia coli* DNA polymerase I. *Journal of Biological Chemistry*. 1979;254(24):12269–12272.

10. Serment-Guerrero J., Bravo-Gomez M. E., Lara-Rivera E., Ruiz-Azuara L. Genotoxic assessment of the copper chelated compounds Casiopeinas: Clues about their mechanisms of action. *Journal of Inorganic Biochemistry*. 2017;166;68–75. https://doi.org/10.1016/ j.jinorgbio.2016.11.007.

11. Bravo-Gómez M. E., Dávila-Manzanilla S., Flood-Garibay J., Muciño-Hernández M. Á., Mendoza Á., García-Ramos J. C., et al. Secondary ligand effects on the cytotoxicity of several Casiopeína's group II compounds. *Journal of the Mexican Chemical Society.* 2012;56(1):85–92.

12. Tabti R., Tounsi N., Gaiddon C.,Bentouhami E., Désaubry L. Progress in copper complexes as anticancer agents. *Medicinal Chemistry*. 2017;7(5):875–879. https:// doi.org/10.4172/2161-0444.1000445.

13. Kletskov A. V., Bumagin N. A., Zubkov F. I., Grudinin D. G., Potkin V. I. Isothiazoles in the design and synthesis of biologically active substances and ligands for metal complexes. *Synthesis*. 2020;52(2):159–188. https://doi.org/10.1055/s-0039-1690688.

14. Eremina J. A., Lider E. V., Sukhikh T. S., Klyushova L. S., Perepechaeva M. L., Sheven' D. G., et al. Water-soluble copper(II) complexes with 4,5-dichloro-isothiazole-3-carboxylic acid and heterocyclic N-donor ligands: Synthesis, crystal structures, cytotoxicity, and DNA binding study. *Inorganica Chimica Acta*. 2020;510. Article number 119778. https://doi. org/10.1016/j.ica.2020.119778.

15. Kaberdin R. V., Potkin V. I. Isothiazoles (1,2-thiazoles): synthesis, properties and applications. *Russian Chemical Reviews*. 2002;71(8):673–694.

16. De Oliveira Silva A., McQuade J., Szostak M. Recent Advances in the Synthesis and Reactivity of Isothiazoles. *Advanced Synthesis and Catalysis*. 2019;361(13):3050–3067. https://doi.org/10.1002/adsc.201900072.

17. Perdew J. P., Burke K., Ernzerhof M. Generalized gradient approximation made simple. *Physical Review Letters.* 1996;77(18):3865–3868. https://doi.org/10.1103/PhysRevLett.77.3865.

18. Van Lenthe E., Baerends E. J. Optimized Slater-type basis sets for the elements 1-118. *Journal of Computational Chemistry*. 2003;24(9):1142–1156. https://doi.org/10.1002/jcc.10255.

19. Sheldrick G. M. SHELXT – Integrated space-group and crystal-structure determination. *Acta Crystallographica, Section A: Foundations of Crystallography.* 2015;A71(1):3–8. https://doi.org/10. 11 07/S2053273314026370.

20. Sheldrick G. M. Crystal structure refinement with SHELXL. *Acta Crystallographica, Section C: Structural Chemistry*. 2015;C71(1):3–8. https://doi. org/10.1107/S2053229614024218 3.

21. Dolomanov O. V., Bourhis L. J., Gildea R. J., Howard J. A. K., Puschmann H. OLEX2: A complete structure solution, refinement and analysis program. *Journal of Applied Crystallography*. 2009;42(2):339– 341. http://dx.doi.org/10.1107/S0021889808042726.

22. Eremina J. A., Lider E. V., Kuratieva N. V., Samsonenko D. G., Klyushova L. S., Sheven' D. G., et al. Synthesis and crystal structures of cytotoxic mixed-ligand copper(II) complexes with alkyl tetrazole and polypyridine derivatives. *Inorganica Chimica Acta*. 2021;516. Article number 120169. https://doi.org/10.1016/j.ica.2020.120169.

23. Pearson R. G. Absolute electronegativity and hardness: applications to organic chemistry. *Journal of Organic Chemistry*. 1989;54(6):1423–1430. https://doi.org/10.1021/jo00267a034.

24. Eremina J. A., Ermakova E. A., Smirnova K. S., Klyushova L. S., Berezin A. S., Sukhikh T. S., et al. Cu(II), Co(II), Mn(II) complexes with 5-phenyl-tetrazole and polypyridyl ligands: Synthesis, characterization and evaluation of the cytotoxicity and antimicrobial activity. *Polyhedron.* 2021;206. Article number 115352. https://doi.org/10.1016/j.poly.2021.115352.

список источников

1. Bencini A., Lippolis V. 1,10-Phenanthroline: A versatile building block for the construction of ligands for various purposes // Coordination Chemistry Reviews. 2010. Vol. 254, no. 17. P. 2096–2180. https://doi.org/10.1016/j.ccr.2010.04.008.

2. Krasnovskaya O., Naumov A., Guk D., Gorel-

kin P., Erofeev A., Beloglazkina E., et al. Copper coordination compounds as biologically active agents // International Journal of Molecular Sciences. 2020. Vol. 21, no. 11. P. 3965. https://doi.org/10.3390/ijms21113965.

3. Dey D., Roy A. B., Ranjani A., Gayathri L.,

Chandraleka S., Dhanasekaran D., et al. Synthesis and bio-catalytic activity of isostructural cobalt(III)phenanthroline complexes // Journal of Chemical Sciences. 2015. Vol. 127, no. 4. P. 649–661. https:// doi.org/10.1007/s12039-015-0817-y.

4. Al-Omair M. A. Biochemical activities and electronic spectra of different cobalt phenanthroline complexes // Arabian Journal of Chemistry. 2019. Vol. 12, no. 7. P. 1061–1069. https://doi.org/10.101 6/j.arabjc.2018.11.006.

5. Čongrádyová A., Jomová K., Kuckova L., Kožíšek J., Moncol' J., Valko M. Antimicrobial activity of copper(II) complexes // Journal of Microbiology, Biotechnology and Food Sciences. 2014. Vol. 3, special issue 1. P. 67–70.

6. Mahalakshmi R., Raman N. A Therapeutic journey of mixed ligand complexes containing 1,10-phenanthroline derivatives: A review // International Journal of Current Pharmaceutical Research. 2016. Vol. 8, no. 3. P. 1–6.

7. Viganor L., Howe O., McCarron P., McCann M., Devereux M. The antibacterial activity of metal complexes containing 1,10-phenanthroline: Potential as alternative therapeutics in the era of antibiotic resistance // Current Topics in Medicinal Chemistry. 2016. Vol. 17, no. 11. P. 1280–1302. https://doi.org/ 10.2174/1568026616666161003143333.

8. Galindo-Murillo R., García-Ramos J. C., Ruiz-Azuara L., Cheatham T. E., Cortés-Guzmán F. Intercalation processes of copper complexes in DNA // Nucleic Acids Research. 2015. Vol. 43, no. 11. P. 5364–5376. https://doi.org/10.1093/nar/gkv467.

9. Sigman D. S., Graham D. R., D'Aurora V., Stern A. M. Oxygen-dependent cleavage of DNA by the 1,10-phenanthroline.cuprous complex. Inhibition of *Escherichia coli* DNA polymerase I // Journal of Biological Chemistry. 1979. Vol. 254, no. 24. P. 12269–12272.

10. Serment-Guerrero J., Bravo-Gomez M. E., Lara-Rivera E., Ruiz-Azuara L. Genotoxic assessment of the copper chelated compounds Casiopeinas: Clues about their mechanisms of action // Journal of Inorganic Biochemistry. 2017. Vol. 166. P. 68–75. https://doi.org/10.1016/j.jinorgbio.2016.11.007.

11. Bravo-Gómez M. E., Dávila-Manzanilla S., Flood-Garibay J., Muciño-Hernández M. Á., Mendoza Á., García-Ramos J. C., et al. Secondary ligand effects on the cytotoxicity of several Casiopeína's group II compounds // Journal of the Mexican Chemical Society. 2012. Vol. 56, no. 1. P. 85–92.

12. Tabti R., Tounsi N., Gaiddon C.,Bentouhami E., Désaubry L. Progress in copper complexes as anticancer agents // Medicinal Chemistry. 2017. Vol. 7, no. 5. P. 875–879. https://doi.org/10.4172/2161-0444.1000445.

13. Kletskov A. V., Bumagin N. A., Zubkov F. I., Grudinin D. G., Potkin V. I. Isothiazoles in the design and synthesis of biologically active substances and ligands for metal complexes // Synthesis. 2020. Vol. 52, no 2. P. 159–188. https://doi.org/10.1055/s-0039-1690688.

14. Eremina J. A., Lider E. V., Sukhikh T. S., Klyushova L. S., Perepechaeva M. L., Sheven' D. G., et al. Water-soluble copper(II) complexes with 4,5-dichloro-isothiazole-3-carboxylic acid and heterocyclic N-donor ligands: Synthesis, crystal structures, cytotoxicity, and DNA binding study // Inorganica Chimica Acta. 2020. Vol. 510. Article number 119778. https://doi.org/10.1016/j.ica.2020.119778.

15. Kaberdin R. V., Potkin V. I. Isothiazoles (1,2-thiazoles): synthesis, properties and applications // Russian Chemical Reviews. 2002. Vol. 71, no. 8. P. 673–694.

16. De Oliveira Silva A., McQuade J., Szostak M. Recent Advances in the Synthesis and Reactivity of Isothiazoles // Advanced Synthesis and Catalysis. 2019. Vol. 361, no. 13. P. 3050–3067. https://doi.org/ 10.1002/adsc.201900072.

17. Perdew J. P., Burke K., Ernzerhof M. Generalized gradient approximation made simple // Physical Review Letters. 1996. Vol. 77, no. 18. P. 3865–3868. https://doi.org/10.1103/PhysRevLett.77.3865.

18. Van Lenthe E., Baerends E. J. Optimized Slater-type basis sets for the elements 1-118 // Journal of Computational Chemistry. 2003. Vol. 24, no. 9. P. 1142–1156. https://doi.org/10.1002/jcc.10255.

19. Sheldrick G. M. SHELXT – Integrated space-group and crystal-structure determination // Acta Crystallographica, Section A: Foundations of Crystallography. 2015. Vol. A71, no. 1. P. 3–8. https://doi.org/10.1107/S2053273314026370.

20. Sheldrick G. M. Crystal structure refinement with SHELXL // Acta Crystallographica, Section C: Structural Chemistry. 2015, vol. C71, no. 1, pp. 3–8. https://doi.org/10.1107/S2053229614024218 3.

21. Dolomanov O. V., Bourhis L. J., Gildea R. J., Howard J. A. K., Puschmann H. OLEX2: A complete structure solution, refinement and analysis program // Journal of Applied Crystallography. 2009. Vol. 42, no 2. P. 339–341. http://dx.doi.org/10.1107/S002188 9808042726.

22. Eremina J. A., Lider E. V., Kuratieva N. V., Samsonenko D. G., Klyushova L. S., Sheven' D. G., et al. Synthesis and crystal structures of cytotoxic mixed-ligand copper(II) complexes with alkyl tetrazole and polypyridine derivatives // Inorganica Chimica Acta. 2021. Vol. 516. Article number 120169. https://doi.org/10.1016/j.ica.2020.120169.

23. Pearson R. G. Absolute electronegativity and hardness: applications to organic chemistry // Journal of Organic Chemistry. 1989. Vol. 54, no. 6. P. 1423–1430. https://doi.org/10.1021/jo00267a034.

24. Eremina J. A., Ermakova E. A., Šmirnova K. S., Klyushova L. S., Berezin A. S., Sukhikh T. S., et al. Cu(II), Co(II), Mn(II) complexes with 5-phenyltetrazole and polypyridyl ligands: Synthesis, characterization and evaluation of the cytotoxicity and antimicrobial activity // Polyhedron. 2021. Vol. 206. Article number 115352. https://doi.org/10.1016/j.poly.2021.115352.

INFORMATION ABOUT THE AUTHORS

Yuliya A. Golubeva,

Postgraduate Student, Nikolaev Institute of Inorganic Chemistry SB RAS, 3, Acad. Lavrentiev Ave., Novosibirsk, 630090, Russian Federation, julia1995@ngs.ru https://orcid.org/0000-0002-5404-5357

Ksenia S. Smirnova,

Postgraduate Student, Nikolaev Institute of Inorganic Chemistry SB RAS. 3, Acad. Lavrentiev Ave., Novosibirsk, 630090, Russian Federation, smirnova_ksenya96@mail.ru https://orcid.org/0000-0002-6345-3467

Lubov' S. Klyushova,

Junior Researcher, Institute of Molecular Biology and Biophysics, Federal Research Center foe Fundamental and Translational Medicine, 2/12, Timakov St., Novosibirsk, 630060, Russian Federation, klyushovals@mail.ru https://orcid.org/0000-0003-4820-2536

Vladimir I. Potkin,

Dr. Sci. (Chemistry), Professor, Institute of Physical Organic Chemistry of the National Academy of Sciences of Belarus, 13, Surganov St., Minsk, 220072, Belarus, potkin@ifoch.bas-net.by https://orcid.org/0000-0001-7823-3208

Elizaveta V. Lider, Cand. Sci. (Chemistry), Senior Scientist, Nikolaev Institute of Inorganic Chemistry SB RAS, 3, Acad. Lavrentiev Ave., Novosibirsk, 630090, Russian Federation, lisalider@ngs.ru https://orcid.org/0000-0003-4363-6829

Contribution of the authors

The authors contributed equally to this article.

Conflict interests

The authors declare no conflict of interests regarding the publication of this article. The final manuscript has been read and approved by all the co-authors.

Information about the article

The article was submitted 19.10.2021. Approved after reviewing 15.11.2021. Accepted for publication 30.11.2021.

СВЕДЕНИЯ ОБ АВТОРАХ

Ю. А. Голубева, аспирант, Институт неорганической химии им. А. В. Николаева СО РАН, 630090, Новосибирск, пр-т Академика Лаврентьева, 3, Российская Федерация, julia1995@ngs.ru https://orcid.org/0000-0002-5404-5357

К.С.Смирнова,

аспирант, Институт неорганической химии им. А.В. Николаева СО РАН, 630090, Новосибирск, пр-т Академика Лаврентьева, 3, Российская Федерация, smirnova_ksenya96@mail.ru https://orcid.org/0000-0002-6345-3467

Л. С. Клюшова.

младший научный сотрудник, Научно-исследовательский институт молекулярной биологии и биофизики структурное подразделение Федерального исследовательского центра фундаментальной и трансляционной медицины, 630060, Новосибирск, ул. Тимакова, 2/12, Российская Федерация, klyushovals@mail.ru https://orcid.org/0000-0003-4820-2536

В. И. Поткин,

д.х.н., профессор. Институт физико-органической химии НАН Беларуси, 220072, Минск, ул. Сурганова, 13, Беларусь, potkin@ifoch.bas-net.by https://orcid.org/0000-0001-7823-3208

Е. В. Лидер.

к.х.н., старший научный сотрудник, Институт неорганической химии им. А. В. Николаева СО РА, 630090, Новосибирск, пр-т Академика Лаврентьева, 3, Российская Федерация, lisalider@ngs.ru https://orcid.org/0000-0003-4363-6829

Вклад авторов

Все авторы сделали эквивалентный вклад в подготовку публикации.

Конфликт интересов

Авторы заявляют об отсутствии конфликта интересов.

Все авторы прочитали и одобрили окончательный вариант рукописи.

Информация о статье*

Поступила в редакцию 19.10.2021. Одобрена после рецензирования 15.11.2021. Принята к публикации 30.11.2021.

https://vuzbiochemi.elpub.ru/jour